1571

(b) With 10% HCl. A suspension of (+)-1 (1.00 g, 4 mmol) in 10% HCl (10 mL) was heated at 90 °C for 2 h. After the suspension was cooled, the resulting precipitates were collected, washed with H₂O, and dried to give racemic 1 (0.97 g, 97%): mp > 280 °C; $[\alpha]^{22}_{D}$ +0.0° (c 1, DMF). **Racemization of** (-)-1. Racemization of (-)-1 (1.00 g, 4 mmol)

Racemization of (-)-1. Racemization of (-)-1 (1.00 g, 4 mmol) was carried out according to method b described for the racemization of (+)-1 to give racemic 1 (0.95 g, 95%): mp > 280 °C; $[\alpha]^{22}_{D}$ -0.1° (c 1, DMF).

Racemization of 6a. Racemization of **6a** (1.00 g, 3.8 mmol) was carried out according to method b described for the racemization of (+)-1 to give racemic **6a** (0.91 g, 91%): mp > 280 °C; $[\alpha]^{22}_{D} + 0.1^{\circ}$ (c 1, DMF).

Racemization of 6b. Racemization of **6b** (1.00 g, 3.4 mmol) was carried out in 10% HCl (10 mL) at 90 °C for 15 min to give racemic **6b** (0.90 g, 90%): mp 249–250 °C dec; $[\alpha]^{26}{}_{\rm D} 0.0^{\circ}$ (c 1, DMF).

Rate Constants for Racemization of (+)-1, 6a, and 6b. A solution of (+)-1, **6a**, or **6b** (20 mg) in 20% HCl (1.5 mL)-DMF (0.5 mL) was stirred at 90 °C (or 50 °C) in a sealed tube. After the solution was cooled, the optical rotation (α_t) of the resulting compound was measured. These racemizations can be regarded as first-order reactions because linear relationships are found between $\ln \alpha_0/\alpha_t$ and time t, as shown in Figure 2. The rate constant of racemization (k_R /h) was calculated by the least-squares method based on eq 1, where α_0 is the initial value of the optical rotation at time t h.

$$\ln \alpha_0 / \alpha_t = k_{\rm R} t \tag{1}$$

Crystallography. The diffraction experiment for the complex of (+)-1 with brucine was carried out using a colorless transparent prism with dimensions of $0.70 \times 0.30 \times 0.10 \text{ mm}^3$ obtained from ethanol-water. The four-circle diffractometer (AFC/5, RIGAKU) was used with graphite-monochromated CuK α radiation ($\lambda =$ 1.5418 Å). The unit cell dimensions were determined from angular setting of 20 reflections (2 θ values in the range of 40–60°). The structure was solved by the direct methods using SIR85¹⁴ and difference Fourier method. The refinement of atomic parameters was carried out using full-matrix least-squares methods with anisotropic temperature factors for the non-hydrogen atoms. Of 36 hydrogen atoms, 23 atoms were located on the difference Fourier maps and refined with isotropic temperature factors. The positions of other hydrogen atoms were assumed geometrically and fixed throughout the refinement. The anomalous scattering factors of N and O atoms were included in the refinement. Throughout the refinement, the function $\sum w(|Fo| - |Fc|)^2$ was minimized. During the final refinement stage, the weighting scheme of $\sqrt{w} = 1/\sigma |Fo|$ was used. The final R value was 0.052 (Rw = 0.061). The atomic scattering factors were taken from International Tables for X-ray Crystallography.¹⁵

Acknowledgment. We are grateful to Dr. I. Chibata, President, and Dr. S. Saito, Research and Development Executive, for their encouragement and interest. Thanks are due to Drs. T. Tosa, M. Takeda, S. Oshiro, I. Inoue, T. Oine, and R. Yoshioka for their valuable comments during this study.

Supplementary Material Available: Crystal data parameters, final atomic coordinates, anisotropic thermal parameters, bond lengths, and bond angles for the complex of (+)-3 with brucine (21 pages). This material is contained in many libraries on microfiche, immediately follows this article in the microfilm version of the journal, and can be ordered from the ACS; see any current masthead page for ordering information.

Formation of 3-[1'-(Dimethylphenylsilyl)ethyl]azetidin-2-ones: Stereocontrolled Formal Approach to (±)-Thienamycin and (±)-β-(Hydroxyalkyl)aspartic Acid Derivatives

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Received August 7, 1991

Reaction between (\pm) - β -(dimethylphenylsilyl)alkanoyl chlorides and imines of glyoxylic esters provided a route to (\pm) -cis-3-[1'-(dimethylphenylsilyl)ethyl]-4-alkoxycarbonyl β -lactams, while addition of the Fleming's silylcuprate reagent to methyl crotonate and further enolate trapping by the above imines furnished the corresponding (\pm) -trans-3-[1'-(dimethylphenylsilyl)ethyl]-4-alkoxycarbonyl β -lactams. These β -lactams, upon appropriate chemical manipulations, provided a stereocontrolled route to (\pm) -thienamycin precursors and (\pm) - β -(hydroxyalkyl)aspartic acid derivatives.

Appropriately substituted monocyclic azetidin-2-ones have wide applicability in the synthesis of β -lactam antibiotics¹ and also in the preparation of many natural products including both α - and β -amino acids.² One example is the β -lactam 1 derived from aspartic acid,³ which upon chemical manipulations affords suitable intermediates for natural product synthesis,⁴ including a variety of

⁽¹⁴⁾ SIR88: A computer program for automatic analysis of phase problems: Giaccovazzo, C.; Cascarano, G. L.; Polidori, G.; Spagna, R.; Viterbo, D. Acta Crystallogr., Sect. A 1982, 38, 663; Ibid. 1987, 43, 22.
(15) Ibers, J. A., Hamilton, W. C., Eds. International Tables for X-ray Crystallography; Kynoch Press: Birmingham, England, 1974; Vol. IV.

⁽¹⁾ For some reviews on β -lactams antibiotics, see: (a) Chemistry and Biology of β -Lactam Antibiotics; Morin, R. B., Gorman, M., Eds.; Academic: New York, 1982; Vol. 1-3. (b) Recent Advances in The Chemistry of β -Lactam Antibiotics; Brown, A. G., Roberts, S. M., Eds.; The Royal Society of Chemistry: Bourlington House, London, 1984.

⁽²⁾ For a recent review, see: Manhas, M. S.; Wagle, D. R.; Chiang, J.; Bose, A. K. Heterocycles 1988, 27, 1755.

⁽³⁾ Zervas, L.; Winitz, M.; Greenstein, J. P. J. Org. Chem. 1957, 22, 1515.

^{(4) (}a) Coppola, G. M.; Schuster, H. F. in Asymmetric Synthesis; John Wiley: New York, 1987; p 204. For some recent examples, see: (b) Baldwin, J. E.; Adlington, R. M.; Collins, D. W.; Schoffeld, C. J. J. Chem. Soc., Chem. Commun. 1990, 720; Tetrahedron 1990, 46, 4733. (c) Thomas, E. J.; Williams, A. C. J. Chem. Soc., Chem. Commun. 1987, 992.
(d) Roe, J. M.; Thomas, E. J. Synlett 1990, 727.

monocyclic β -lactams like 2, a precursor of the carbapenem antibiotic thienamycin⁵ (3). Aspartic acid itself can also



be elaborated through its β -anion derivative⁶ to nonproteinogenic amino acids, a class of compounds that represents an important group of natural products because of their varied biological and biochemical properties.⁷ For example, β , γ -unsaturated amino acids have been found to be reversible or irreversible enzyme inhibitors⁸ and thus a number of methods have been proposed for their synthesis.^{7c,6d} One of them involves prior formation of β -(hydroxyalkyl)aspartic acids followed by a carboxy-hydroxy elimination step to furnish the desired β , γ -unsaturated amino acids in good to excellent yields.^{6d}

In a recent project directed toward the synthesis of these compounds, we focused our attention on β -lactams 6, which, upon N-C₂ bond cleavage (Scheme I), would render the corresponding aspartic acid derivatives 5 as precursors of β -(hydroxyalkyl)aspartic acids 4. The synthesis of β lactams carrying the 1'-(dimethylphenylsilyl)alkyl group at the C₃ position has been previously reported by Fleming and Kilburn, using the hydroxamate methodology,⁹ and also by Hart et al.¹⁰ through the lithium ester enolateimine condensation. However, condensation of lithium ester enolates with imines of glyoxylic esters has proved to be unsuccessful.¹¹ In general, the metal enolate-glyoxylic ester imine condensation works well with tin(II) enolates, silyl ketene acetals, and boron enolates, to give the corresponding β -substituted aspartic acid derivatives in good to excellent stereoselectivities,¹² which at once could be cyclized by standard procedures.^{12,13} Recently, we have described¹⁴ that the organocopper enolate-glyoxylic ester imine condensation could be an efficient alternative to the above approaches leading to trans β -lac-



a R1: PhMe2Si, b R1: Ph2MeSi, c R1: Ph3Si, d R1: Ph2BuSi

^aReagents and conditions: (i) (R₁)₂CuCNLi₂, THF, 0 ^oC, 20 min; (ii) KOH, MeOH, reflux; (iii) ClCOCOCl, CH₂Cl₂, rt, 1 h.



a R1: PhMe2Si, b R1: Ph2MeSi, c R1: Ph3Si, d R1: Ph2BuSi

 aReagents and conditions: (i) $NEt_3,$ solvent, reflux, 14 h. Abbreviations: PMP, 4-MeOC_6H_4.

tams 6 in both excellent chemical yield and stereoselectivity. Alternatively, the β -(dimethylphenylsilyl)butanoyl chloride-glyoxylic ester imine condensation¹⁵ was found to be a good approach for synthesis of the corresponding cis β -lactams 6.

In this paper we present the details of our methodology for the construction of both cis and trans β -lactams of type 6 as well as their utility in the synthesis of the (\pm) -thienamycin precursor 7 (R = Me) and (\pm) - β -(hydroxyalkyl)aspartic acid derivatives of type 4 with control of the relative stereochemistry at the three chiral centers.

Results and Discussion

Acid Chloride-Methyl Glyoxalate Imine Condensation. The construction of monocyclic β -lactams by the [2 + 2] cycloaddition reaction of ketenes to imines has proved to be particularly successful with acid chlorides bearing electron-withdrawing substituents at the α -position. However, the direct preparation of 3-alkyl β -lactams from monoalkylketenes, generated from their corresponding acid chlorides, is often limited in scope.¹⁶ Even if the formation of 3-(1'-hydroxyethyl) β -lactams has been reported¹⁷ to occur from O-protected 3-hydroxybutyric acid chlorides and phenyl glyoxal imines, such a reaction affords only low yields (3-10%) of β -lactams when carried out

^{(5) (}a) Salzmann, T. N.; Ratcliffe, R.; Christensen, B. G.; Bouffard, F. A. J. Am. Chem. Soc. 1980, 102, 6161. (b) Reider, P. J.; Grabowski, E. J. J. Tetrahedron Lett. 1982, 23, 2293. (c) 1-β-Methylthienamycin: Rama Rao, A. V.; Gurjar, M. K.; Khare, V. B.; Ashok, B.; Deshmukh, M. N. Tetrahedron Lett. 1990, 31, 271. For a recent review on thienamycin, see: Georg, G. I. In Studies in Natural Product Chemistry; Rahman, A-ur, Ed.; Elsevier: Amsterdam, 1989; Vol. 4, p 431.

<sup>Georg, G. I. In Studies in Natural Product Chemistry; Rahman, A-ur, Ed.; Elsevier: Amsterdam, 1989; Vol. 4, p 431.
(6) (a) Wolf, J. P.; Rapoport, H. J. Org. Chem. 1989, 54, 3164. (b) Seebach, D.; Wasmuth, D. Angew. Chem., Int. Ed. Engl. 1981, 20, 971.
(c) Baldwin, J. E.; Moloney, M. G.; North, M. Tetrahedron 1989, 45, 6309.
(d) Baldwin, J. E.; Moloney, M. G.; North, M. Tetrahedron 1989, 45, 6319.</sup>

^{(7) (}a) Chemistry and Biochemistry of the Amino Acids; Barrett, G. C., Ed.; Chapman and Hall: London, 1985. (b) Wagner, I.; Musso, H. Angew. Chem., Int. Ed. Engl. 1983, 22, 816. (c) Williams, R. M. In Synthesis of Optically Active α-Amino Acids; Pergamon Press: Oxford, 1989.

^{(8) (}a) Rando, R. R. Science 1974, 185, 320. (b) Walsh, C. Tetrahedron
1982, 38, 871. (c) Baldwin, J. E.; Adlington, R. M.; O'Niel, I. A.; Schofield,
C.; Spivey, A. C.; Sweeney, J. B. J. Chem. Soc., Chem. Commun. 1989,
1852. (d) Baldwin, J. E.; Adlington, R. M.; Robinson, N. G. J. Chem. Soc.,
Chem. Commun. 1987, 153. (e) Castelhano, A. L.; Horne, S.; Taylor, G.
J.; Billedeau, R.; Krantz, A. Tetrahedron 1988, 44, 5451. (f) Sibi, M. P.;
Renhowe, P. A. Tetrahedron Lett. 1990, 31, 7407.
(9) Fleming, I.; Kilburn, J. D. J. Chem. Soc., Chem. Commun. 1986,

⁽⁹⁾ Fleming, I.; Kilburn, J. D. J. Chem. Soc., Chem. Commun. 1986, 1198.

⁽¹⁰⁾ Burnett, D. A.; Galluci, J. C.; Hart, D. J. J. Org. Chem. 1985, 50, 5120.
(11) Georg, G. I.; Kant, J.; Gill, H. S. J. Am. Chem. Soc. 1987, 109,

⁽¹¹⁾ Georg, G. I.; Kant, J.; Gill, H. S. J. Am. Chem. Soc. 1987, 109, 1129.

⁽¹²⁾ For recent reviews on ester enolate-imine condensation, see: (a) Brown, M. J. Heterocycles 1989, 29, 2225. (b) Hart, D. J.; Ha, D. C. Chem. Rev. 1989, 89, 1447.

⁽¹³⁾ Palomo, C. In Recent Progress in The Chemical Synthesis of Antibiotics; Lukacs, G., Ohno, H., Eds.; Springer-Verlag Berlin: Berlin, 1990; p 565.

⁽¹⁴⁾ Palomo, C.; Aizpurua, J. M.; Urchegui, R. J. Chem. Soc., Chem. Commun. 1990, 1390.

⁽¹⁵⁾ Palomo, C.; Ontoria, J. M.; Odriozola, J. M.; Aizpurua, J. M.; Ganboa, I. J. Chem. Soc., Chem. Commun. 1990, 248.

⁽¹⁶⁾ For a discussion and references, see: Palomo, C.; Cossio, F. P.; Odriozola, J. M.; Oiarbide, M.; Ontoria, J. M. J. Org. Chem. 1991, 56, 4418.

⁽¹⁷⁾ Lynch, J. E.; Riseman, S. M.; Laswell, W. L.; Tschaen, D. M.; Volante, R. P.; Smith, G. B.; Shinkai, I. J. Org. Chem. 1989, 54, 3792.



^aReagents and conditions: (i) HBF₄·OEt₂, CH₂Cl₂, 0 °C \rightarrow rt, 1 h; (ii) MeCO₃H, AcOH (32%), NEt₃, 0 °C \rightarrow rt, 3 h; (iii) BF₃·OEt₂, CH₂Cl₂, rt, 20 min; (iv) MeSO₂Cl, pyridine, rt, 30 min, then DBU, benzene, reflux.

from glyoxylic ester imines.¹⁸ In previous papers from our^{15,19} and other laboratories,²⁰ it has been established that the reaction of α -unactivated alkanoyl halides with imines of glyoxylic esters efficiently affords the corresponding 3-alkyl β -lactams in a single synthetic step, thus providing a solution to the aforementioned problem. In our approach, several alkanoyl halides can be used as starting materials, most notably, the β -(dimethylphenylsilyl)butanoyl chloride, which, upon treatment with an imine of glyoxylic methyl ester and further N-deprotection of the resulting β -lactam, provided a route to compounds of type 6.

The starting acid chlorides 12 (Scheme II) could be easily prepared by saponification of the corresponding methyl esters and further oxalyl chloride activation of the resulting carboxylic acids. In general, β -silylbutanoic esters can be formed from α,β -unsaturated esters in yields ranging from 50 to 67% by addition of the corresponding silylcuprate reagent to methyl crotonate (8) following a standard protocol.²¹ When the acid chloride 12a (Scheme III) was allowed to react with the methyl glyoxalate imine 14 in refluxing hexane for 14 h, a mixture of cis and trans β -lactams was produced in 76% isolated yield and in a 66:34 ratio, respectively. The cis isomers were isolated from the corresponding trans isomers by column chromatography and the corresponding cis,anti and cis.syn isomers of the β -lactam 15a were separated by crystallization from methanol. In a subsequent experiment to achieve a better stereoselective control of the β -lactam formation, following observations on related [2 + 2] cycloadditions,²² we examined the influence of the solvent. We found that the ratio of cis-anti isomer was increased when the reaction was carried out in a more polar solvent, like acetonitrile. Under these conditions, the ratio of cis isomers increased up to 82% and 18% of the corresponding trans isomers was detected by ¹H NMR from the crude reaction mixture. The relative stereochemistry of diastereomeric β -lactams 15a, anti and syn, respectively, according to the nomenclature introduced by Masamune,²³ was established on the basis of their respective ¹H NMR spectra and some chemical correlations which will be discussed later.

After studying the influence of the solvent on the stereoselectivity of the reaction, the cycloaddition was examined for different β -silylalkanoyl chlorides carrying diverse silyl groups. Thus, under the same conditions as those used for the formation of 15a, compounds 15b-d were obtained in yields varying in the range 50-86%. The results summarized in Table I indicated that an increase in the bulkiness of the silyl moiety has little effect on the cis:trans ratio, but a moderate increase in the anti:syn ratio is observed for cis isomers by using β -(triphenylsilyl)butanoyl chloride (12c) or β -(tert-butyldiphenylsilyl)butanoyl chloride (12d). Furthermore, the cis-anti isomers of compounds 15c and 15d were readily isolated from the reaction crudes by crystallization from methanol, affording these compounds in 41% and 60% isolated yields. The stereochemistry of cis and trans isomers was determined on the basis of their ¹H NMR spectral data. Thus, the proton at C_4 in cis isomers shows as a doublet between 4.4 and 4.6 ppm ($J \approx 5$ –6 Hz) while the corresponding one in trans isomers appears at 4.0-4.1 ppm ($J \approx 2$ Hz). In general, the C_4 -H proton in a trans isomer appears at higher fields than the corresponding C_4 -H proton in the respective cis isomer.

The first attempt at determining the ratio of epimers for compounds 15 was (Scheme IV) by the conversion of the silyl moiety into the hydroxyl group and further elimination to the alkenes 20, according to the Pecquet and d'Angelo protocol used on similar compounds.²⁴ Conversion of *anti*-15a into the β -lactam *anti*-17 was easily accomplished following Fleming's methodology.^{9,21} Ac-

⁽¹⁸⁾ Ernst, B.; Bellus, D. D.E. 3620467 A1, 1987; Chem. Abstr. 1987, 106, 176045q.

⁽¹⁹⁾ Palomo, C.; Cossio, F. P.; Ontoria, J. M.; Odriozola, J. M. J. Org. Chem. 1991, 56, 5948.

⁽²⁰⁾ Firestone, R. A.; Barker, P. L.; Pisano, J. M.; Ashe, B. M.; Dahlgren, M. E. Tetrahedron 1990, 46, 2255.

⁽²¹⁾ Fleming, I.; Hill, J. H. M.; Parker, D.; Waterson, J. J. Chem. Soc., Chem. Commun. 1985, 318. For a review, see: Fleming, I. Pure Appl. Chem. 1988, 60, 71.

⁽²²⁾ Brady, W. T.; Roe, R., Jr. J. Am. Chem. Soc. 1970, 92, 4618.

⁽²³⁾ Masamune, S.; Kaiho, T.; Garvey, D. S. J. Am. Chem. Soc. 1982, 104, 5521. For another nomenclature for β -lactams, see: Georg, G. I.; Akgun, E. Tetrahedron Lett. 1990, 31, 3267.

⁽²⁴⁾ Pecquet, F.; d'Angelo, J. Tetrahedron Lett. 1982, 23, 2777.

Table I. Results of Cycloaddition Reaction between β -(Trialkylsilyl)alkanoyl Halides 12 and the Methyl Glyoxalate Imine 14^a

compd				ratio isomers 15 ^c				
		solvent	yield, % ^b		trans			
	R_1			anti	mp, °C ^d	syn		
15 a	PhMe ₂ Si	hexane	76	38	107-108	28	34	
	-	CH ₃ CN	47	54	(hexane)	28	18	
15b	Ph ₂ MeSi	hexane	68	37	syrup	32	31	
	2	THF	63	44		34	34 22	
		CH ₃ CN	50	55		27	18	
15c	Ph_3Si	hexane	77	52	224 - 225	23	25	
	Ũ	CH ₃ CN	64	65	(MeOH)	18	17	
15 d	Ph ₂ ^t BuSi	hexane	78	52	168-169	15	33	
	2	C_6H_6	81	58	(hexane– CH ₂ Cl ₂)	14	28	
		CH_3CN	86	70	2 - 2,	14	16	

^aReactions conducted on 5-mmol scale. ^b Yields based on weight of isolated product by column chromatography. ^c All percentages refer to diastereomeric ratios, determined from the crude reaction mixtures. ^d Melting points correspond to *cis,anti*-15 isomers. Crystallization solvent indicated in parentheses.

cordingly, reaction between anti-15a and the $HBF_4 \cdot OEt_2$ complex followed by peracetic acid oxidation of the resulting intermediate fluoride anti-16 afforded the expected hydroxy compound anti-17 in 40% isolated yield together with the cyclized product 18 in 55% yield. Treatment of compound anti-17 with boron trifluoride in methylene chloride as solvent cleanly afforded the lactone 18 in 70% isolated yield. Similarly, when the β -lactam syn-15a was subjected to the foregoing reaction sequence, the hydroxy compound syn-17 was obtained as single reaction product. This compound upon treatment with boron trifluoride provided the expected lactone 19 in 70% isolated yield. Mesylation of both anti and syn isomers of the β -lactam 17 followed by 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU) promoted elimination gave a mixture of the corresponding E and Z alkenes 20. However, from this method it was not possible to determine the relative stereochemistry at $C_{1'}$ and C_3 positions. Under the above conditions, the hydroxy derivative anti-17 afforded E and Z alkenes 20 in a ratio of 2:1, respectively, and the syn isomer of 17 furnished an equimolar mixture of both alkenes. These alkenes were separated by crystallization from ethanol and characterized as previously reported from this laboratory.¹⁶

At this stage, we were able to determine the stereochemistry at $C_{1'}$ and C_3 by means of NOE experiments made on lactones 18 and 19, respectively. Thus, by presaturation of the methyl group of the lactone 19, a 10% NOE was observed in the signal corresponding to the C_3 -H proton, whereas in the case of lactone 18 no enhancement was detected. Consequently, from this observation, we assigned the relative stereochemistry $1'R^*, 3S^*, 4S^*$ to compound anti-15a and $1'R^*, 3R^*, 4R^*$ to compound syn-15a. The most relevant ¹H NMR data of β -lactams 15a-d are compiled in Table II and show that in each case the lower field H₃ and higher field H₄ protons could be assigned to the cis, syn-15 isomers having the larger $J_{3,4}$ values (6.3 Hz) and the smaller $J_{1',3}$ values (2.1-3.6 Hz).²⁵

As the best stereoselection in β -lactam formation was observed for azetidinones 15c and 15d carrying the triphenylsilyl and *tert*-butyldiphenylsilyl groups, respectively, we tried at this stage to achieve their two-step hydroxy-

Table II. Significative ¹H NMR Data ofCis Compounds 15a-da

	δ (ppm)		J (Hz)		
compd	H ₃	H ₄	$\overline{J_{3,4}}$	$J_{1',3}$	
anti-15a	3.41	4.54	5.7	12.3	
syn-15a	3.70	4.43	6.3	3.6	
anti-15b	3.46	4.54	5.7	11.7	
syn-15b	3.71	4.42	6.3	3.0	
anti-15c	3.51	4.57	5.7	11.1	
syn-15c	4.07	4.45	6.3	2.7	
anti-15d	3.42	4.65	5.7	9.6	
syn-15 d	3.96	4.42	6.3	2.1	

 $^{\rm a} \rm Determined$ by 300-MHz $^1\rm H$ NMR spectroscopy in $\rm CDCl_3$ solution.



^aReagents and conditions: (i) NEt₃, solvent, reflux, 14 h. Abbreviations: DAM, $(4-MeOC_6H_4)_2CH$.

lation. However, treatment of the triphenylsilyl β -lactam 15c with the HBF_4 ·OEt₂ complex followed by peracetic acid oxidation under Fleming's protocol did not allow the formation of any detectable amount of the expected 3-(1'-hydroxyethyl) β -lactam. In contrast, the parent *tert*butyldiphenylsilyl derivative 15d upon treatment with the $HBF_4 \cdot OEt_2$ complex in methylene chloride containing acetic acid afforded, after 20 h of reaction at room temperature, the desired fluorosilane. Unfortunately, attempted transformation of such a fluorosilane into the corresponding hydroxy compound by means of peracetic acid was unfruitful and the starting compound was recovered unchanged. The use of other oxidizing reagents, such as *m*-chloroperbenzoic acid-triethylamine²⁶ or KF/ H_2O_2 in N,N-dimethylformamide,²⁷ caused formation of complex mixtures without detection of the desired 3-(1'hydroxyethyl) β -lactam 17.

In view of the preceding results, the dimethylphenylsilyl group was considered the most adequate to constitute a precursor of the hydroxy function in β -lactams. Consequently, we next attempted to improve the stereoselectivity of the cycloaddition step by increasing the bulkyness of the imino component of the reaction.²⁸ To achieve our goal (Scheme V), we examined the method for the imine 21, derived from methyl glyoxalate and di-*p*-anisylmethylamine (DAM-NH₂).²⁹ We have found that when a mixture of the methyl glyoxalate imine 21 and triethylamine was treated with β -(dimethylphenylsilyl)butanoyl chloride (12a) in refluxing hexane for 14 h, an 88% yield of the corresponding cis isomers of the β -lactam 22 was obtained without traces of the corresponding trans isomers. A similar result was found when the acid chloride

^{(25) (}a) For related observations on similar compounds, see: Burnett, D. A.; Gallucci, J. G.; Hart, D. J. J. Org. Chem. 1985, 50, 5120. (b) For correlation data on 3-(1'-hydroxyethyl) β -lactams, see: Cainelly, G.; Panuncio, M.; Basile, T.; Bongini, A.; Giacomini, O.; Martelli, G. J. Chem. Soc., Perkin Trans. I 1987, 2637.

⁽²⁶⁾ Tamao, K.; Kakui, T.; Kumada M. J. Am. Chem. Soc. 1978, 100, 2268.

⁽²⁷⁾ Tamao, K.; Ishida, N. Tetrahedron Lett. 1984, 25, 4249.

 ^{(28) (}a) Moore, H. W.; Hughes, G.; Srinivasachar, K.; Fernandez, M.;
 Nguyen, N. V.; Schau, D.; Tranne, A. J. J. Org. Chem. 1985, 50, 4231. (b)
 Arrieta, A.; Lecea, B.; Palomo, C. J. Chem. Soc., Perkin Trans. I 1987,
 845. (c) Aizpurua, J. M.; Cossio, F. P.; Lecea, B.; Palomo, C. Tetrahedron Lett. 1986, 27, 4359.

^{(29) (}a) Kobayashi, Y.; Ito, Y.; Terashima, S. Bull. Chem. Soc. Jpn. (29) (a) Kobayashi, Y.; Ito, Y.; Terashima, S.; Sasaki, A.; Sunagawa, M. Tetrahedron 1988, 44, 2149. (c) Ito, Y.; Kobayashi, Y.; Kawabata, T.; Takase, M.; Terashima, S. Tetrahedron 1989, 45, 5767.

Table III. Results of Cycloaddition Reaction between β -(Trialkylsilyl)alkanoyl Halides 12a and 13a and Methyl Glyoxalate Imine 21^a

compd		solvent		isomers vield. ratio ^c		¹ H NMR			
			vield.			anti		syn	
	R		% ^b	anti s	syn	$\delta(H_4)$	$J_{1',3}$	$\overline{\delta(H_4)}$	$J_{1',3}$
22	Me	hexane	88	66	34	4.03	12.3	3.88	3.7
		CH ₂ Cl ₂	76	70	30				
		CH ₃ CN	70	71	29				
23	Ph	CH ₃ CN	70	68	32	3.80	12.9	3.74	4.0
		CH ₂ Cl ₂	71	91	9				

^aReactions conducted on 5-mmol scale. ^bYields based on weight of isolated product by column chromatography. ^cAll percentages refer to diastereomeric ratios, determined from the crude reaction mixtures.

Table IV. Preparation of β -Lactams 29–30 from α,β -Unsaturated Esters 27 and Methyl Glyoxalate Imine 14^{α}

	vield.		¹ H NMR				
compd	% ^b	mp, °C	$\delta(H_4)$	$\delta(\mathbf{H}_3)$	$J_{3,4}$	$J_{1',3}$	
29a	80	80-82	4.16	3.34	2.4	5.1	
29b	65	syrup	3.91	2.81	2.4	10.5	
30b	11	syrup	4.37	4.10	5.7	13.5	
29c	85	89-90	4.19	3.25	2.5		
29d	74	syrup	3.91	3.75	2.4	10.2	
30d	64	syrup	4.32	4.06	5.5	13.4	

^a Reactions conducted on 5-mmol scale. ^b Yields based on weight of isolated product by column chromatography.

13a was employed in such a cycloaddition reaction to furnish the desired β -lactam 23 in 71% yield exclusively as the cis isomer. Results of some experiments are listed in Table III to illustrate the efficiency of this bulky Schiff base to control the stereoselectivity of the cycloaddition reaction.

Organocopper Enolate-Glyoxylic Ester Imine Condensation. In a previous paper from our laboratory,¹⁶ we showed that addition of Fleming's silylcuprate reagent to α -ethylidene β -lactams provided a route to 3-[1'-(dimethylphenylsilyl)ethyl] β -lactams in good yields but in modest stereoselectivity. The straightforward methodology for the conjugate addition of a silylcuprate reagent to α , β -unsaturated esters 24 followed by enolate trapping by



carbonyl compounds constitutes an excellent general procedure for the synthesis of aldol products 25 with concomitant construction of three constiguous chiral centers under very high levels of stereoselectivity.^{9,21} Based on this principle, we thought that the use of a methyl glyoxalate imine instead of a carbonyl compound in the last step of the above protocol could give the β -amino ester 26 with an analogous level of stereocontrol. We found that addition of Fleming's silylcuprate reagent to methyl crotonate (27a), followed by trapping of the in situ-generated enolate by the methyl glyoxalate imine 14 in THF as solvent, afforded directly the β -lactam 29 in 80% yield as the single diastereomer (Scheme VI). Similarly (see Table IV), when the above two-step procedure was carried out on methyl cinnamates 27b and 27d, the corresponding trans β -lactams 29b and 29d were produced as major diastereomers together with small amounts of the corresponding cis isomers 30. In the case of methyl 2methylbut-2-enoate (27c), only the trans β -lactam 29c was

Scheme VI^a







^aReagents and conditions: (i) (PhMe₂Si)₂CuCNLi₂, THF, 0 °C, 20 min; (ii) PMP-N=CHCO₂Me (14), THF, $0 \rightarrow 20$ °C, 3 h.



[°]Reagents and conditions: (i) HBF₄·OEt₂, CH₂Cl₂, 0 °C \rightarrow 20 h; (ii) MeCO₃H, MeCO₂H (32%), NEt₃, 0 °C \rightarrow rt, 3 h; (iii) NDC, pyridine, C₆H₆, rt 2 h; (iv) MeSO₂Cl, pyridine, rt, 30 min, then DBU, C₆H₆, reflux; (v) PPh₃, DEAD, HCO₂H, 0 \rightarrow 25 °C, 1.5 h, then MeOH, HCl, 25 °C, 1 h; (vi) NaBH₄, MeOH-THF (2:1), rt, 30 min.

produced in 85% yield. The assignment of a cis or a trans stereochemistry to these compounds was made by examining the values of the coupling constants $J_{3,4}$, vide supra.

On the other hand, the relative stereochemistry of compound **29a** at $C_{1'}$ and C_3 was determined by two different paths (Scheme VII). First, it was submitted to the fluorination-oxidation sequence described earlier for cis analogues, to give the pure 3-(1'-hydroxyethyl) β -lactam **33** in 81% overall yield. Its conversion into the corresponding methanesulfonate, followed by stereoselective elimination,²⁴ afforded the alkene¹⁶ (Z)-20, indicating an anti relationship at $C_{1'}$ - C_3 for the parent compound **29a**. As expected, alkene (E)-20 could also be obtained in a similar fashion from the epimer **35**, which, in turn, was derived quantitatively from **33** by the Mitsunobu reaction³⁰ using

⁽³⁰⁾ Mitsunobu, O. Synthesis 1981, 1. For application in β -lactam chemistry, see, for example: Corbett, D. F.; Coulton, S.; Southgate, R. J. Chem. Soc., Perkin Trans. 1 1982, 3011. Melillo, D. G.; Shinkai, T.; Liu, K.; Ryan, K.; Sletzinger, M. Tetrahedron Lett. 1980, 21, 2783.



^aReagents and conditions: (i) $(NH_4)_2Ce(NO_3)_6$, $CH_3CN\cdot H_2O$, -10 °C, 3 h; (ii) $ClSiMe_3$ (5 equiv), MeOH, rt, 4 h; (iii) PhCOCl, NEt₃, CH_2Cl_2 , rt; (iv) $HBF_4\cdot OEt_2$, CH_2Cl_2 , 0 °C \rightarrow rt, 16 h; (v) CH_3CO_3H (32% in AcOH), NEt₃, CH_2Cl_2 , 0 °C \rightarrow rt, 3.5 h.

formic acid as a nucleophile, followed by acid hydrolysis. The second path for confirming the proposed stereochemistry consisted of the borohydride reduction of the 3-acetyl β -lactam 37, easily obtained by oxidation³¹ of 33. The result of such a reduction was a 60:40 mixture of alcohols 33 and 35 in which the anti epimer predominates, in good agreement with previous observations of Bouffard el at. on related reductions.³² This last method was choosen to confirm the relative stereochemistry at $C_1 - C_3$ for the β -lactam 29b. Thus, the same sequence of reactions as above gave the expected 3-benzoyl β -lactam 38 in good yield. Reduction of this compound with sodium borohydride in methanol resulted in an epimeric mixture of alcohols (75:25) in which the major isomer was assigned to the anti epimer 34.

Since 35 can be stereospecifically obtained from 29a via Mitsunobu reaction and both N_1 and C_4 groups can be easily elaborated to the β -lactam 2,^{5,11} our procedure constitutes a highly stereoselective formal synthesis of (±)-thienamycin (3).

Formation of β -(Hydroxyalkyl)aspartic Acids. Conversion of the β -lactams, prepared as above, into β -(hydroxyalkyl)aspartic acids was accomplished following the sequences illustrated in Schemes VIII and IX. First, the cis β -lactam anti-22 was N-deprotected³³ (Scheme VIII), giving the expected NH azetidin-2-one **39** in 80% yield. The β -lactam ring opening, achieved by means of trimethylchlorosilane in methanol,³⁴ afforded the expected dimethyl aspartate 40a in 70% yield, which was isolated as the N-benzoyl derivative 41a. Similarly, the β -lactam anti-23 upon N-deprotection and further β -lactam cleavage of the resulting NH azetidin-2-one **39b** furnished the β amino ester 40b in 83% yield, which was also isolated as the N-benzoyl derivative 41b. Conversion of these compounds into their corresponding hydroxy derivatives was performed according to the usual Fleming protocol. For



^a Reagents and conditions: (i) $(NH_4)_2Ce(NO_3)_6$, $CH_3CN\cdot H_2O$, 0 °C, 45 min; (ii) $ClSiMe_3$ (5 equiv), MeOH, rt, or reflux; (iii) PhCOCl, NEt₃, CH_2Cl_2 , rt; (iv) $HBF_4\cdot OEt_2$, CH_2Cl_2 , 0 °C \rightarrow rt, 16 h; (v) CH_3CO_3H (32% in AcOH), NEt₃, CH_2Cl_2 , 0 °C \rightarrow rt, 3.5 h.

example, when compound 41a was successively treated with the HBF₄·OEt₂ complex and peracetic acid, the corresponding dimethyl β -(hydroxyalkyl)aspartate was obtained in 72% yield. Under similar reaction conditions 41b provided 42b in 65% isolated yield.

Next, the above sequence of reactions was tested for trans 4-methoxycarbonyl β -lactams 29, Scheme IX, prepared by the organocopper enolate-imine methodology. Thus, the β -lactam 29a upon N-dearylation furnished the NH azetidin-2-one 43a in 92% yield, which was isolated by column chromatography, using hexane-EtOAc (1:1) as eluant. Similarly, compounds 29b and 29c gave the corresponding N-unsubstituted derivatives 43 in yields of 70% and 65%, respectively. The opening of the β -lactam ring was achieved as above by treating compounds 43a and 43b with trimethylchlorosilane-methanol at room temperature. In the case of compound 43c, the reaction might be accomplished under reflux conditions to give 44c in 95% yield. All of these compounds were obtained as syrups and used without further purification in the next step. Protection of the amino group as the benzoyl derivative was carried out under usual conditions by using a slight excess of benzoyl chloride and triethylamine as base and the resulting products were purified by flash column chromatography using hexane-methylene chloride (15:1) as eluant to separate the remaining benzoyl chloride.

Conversion of these β -amino esters into the corresponding β -(hydroxyalkyl)aspartates 46 was achieved under standard conditions by treating them with the HBF₄-OEt₂ complex in methylene chloride at room temperature. Compound 45a was completely transformed into its corresponding fluoride with 3 equiv of the above complex, but compounds 45b and 45c required 5 equiv of the reagent for completion. The resulting crude fluorides were subjected to treatment with peracetic acid in the presence of triethylamine to furnish the dimethyl β -(hydroxyalkyl)aspartates 46a-c in excellent yields.

Conclusion

In summary, the use of the dimethylphenylsilyl group as a masked hydroxy function and the choice of the appropriate method for the azetidin-2-one ring formation (acid chloride or organocopper enolate) allows a highly diastereoselective transformation of glyoxylic ester derived imines into either *cis*- or *trans*-3-[1'-(dimethylphenylsilyl)alkyl]-4-methoxycarbonyl β -lactams as intermediates for both β -lactam antibiotics and β -(hydroxyalkyl)aspartic

⁽³¹⁾ Cossio, F. P.; Lôpez, M. C.; Palomo, C. Tetrahedron 1987, 43, 3963. See also: Matikainen, K. T.; Kaltia, S. A. A.; Hase, T. A.; Ssundberg, M. R. J. Chem. Res., Synop. 1990, 150; J. Chem. Res., Miniprint 1990, 1117.

 ⁽³²⁾ Bouffard, F. A.; Christensen, B. G. J. Org. Chem. 1981, 46, 2208.
 (33) Kronenthal, D. R.; Han, C. Y.; Taylor, M. K. J. Org. Chem. 1982, 47, 2765.

⁽³⁴⁾ Palomo, C.; Arrieta, A.; Cossio, F. P.; Aizpurua, J. M.; Mielgo, A.; Aurrekoetxea, N. *Tetrahedron Lett.* 1990, 31, 6429. For related methods, see: (a) Hauser, F. M.; Ellenberger, S. R.; Rhee, R. P. J. Org. Chem. 1987, 52, 5041. (b) For a review on β -lactam cleavage, see: Manhas, M. S.; Wagle, D. R.; Chiang, J.; Bose, A. K. *Heterocycles* 1988, 27, 1755.

acid derivatives. The present study establishes the basis for the preparation of the above compounds in a optically active pure form, via enantiomerically pure β -silyl carboxylic acids or derivatives.³⁵

Experimental Section

Melting points were determined on a Büchi SMP-20 instrument and are uncorrected. Proton nuclear magnetic resonance spectra and ¹³C NMR spectra were recorded on a Varian VXR 300 spectrometer. All chemical shifts are reported as δ values (ppm) relative to internal tetramethylsilane. Infrared (IR) spectra were recorded on a Shimadzu IR-435 spectrometer. Mass spectra were obtained on a Hewlett-Packard 5930 A spectrometer operated at 70 eV. Microanalytical data were obtained on a Perkin-Elmer 240-C instrument. Commercially available compounds were used in this work without further purification or were prepared by following literature procedures. Acetonitrile and hexane were dried and purified by distillation. Tetrahydrofuran was distilled over sodium and benzophenone (indicator). Methylene chloride was shaken with concentrated sulfuric acid, dried over potassium carbonate, and distilled.

Preparation of β -Silylbutanoic Acids 10a-d and β -Phenyl-*B*-silylpropanoic Acids 11a-c. General Procedure. A solution of the corresponding silvilithium (1 M, 100 mL, 100 mmol) in tetrahydrofuran was added dropwise over dry CuCN (4.46 g. 50 mmol) kept under nitrogen while being stirred at 0 °C. The dark suspension was stirred for 30 min at the same temperature and then methyl crotonate (5.30 mL, 50 mmol) in tetrahydrofuran (5 mL) or methyl cinnamate (8.10 g, 50 mmol) in the same solvent (5 mL) was added. The solution was stirred for 30 min at 0 °C, poured into methylene chloride (20 mL) and saturated NH₄Cl (100 mL), and stirred for 15 min again. Then, the organic layer was filtered through a pad of Celite, dried, and evaporated to yield the corresponding crude methyl β -silylbutanoate, which was saponified without further purification. Thus, KOH (2.8 g, 50 mmol) was added to a solution of the ester in methanol (50 mL) and water (4 mL), and the solution was refluxed for 3 h. This solution was cooled and poured into 0.5 M KOH (200 mL) and diethyl ether (50 mL). The organic layer was separated, and the aqueous phase was washed with diethyl ether (50 mL). The aqueous phase was acidified with 6 N HCl and extracted with diethyl ether (3×50) mL), and the organic layer was dried (Na_2SO_4) and evaporated to yield the corresponding β -silylbutanoic acid, which was purified by reduced pressure distillation or crystallization (hexane/ methylene chloride).

Reaction between β -Silylalkanoyl Chlorides and Methyl Glyoxate Imines. General Procedure for the Preparation of β-Lactams 15, 22, and 23. Oxalyl chloride (0.51 mL, 6 mmol) was added dropwise in methylene chloride (5 mL) to a cold (0 °C) solution of pure β -silvlbutanoic acid or β -phenyl- β -silvlpropanoic acid (3 mmol) in dry methylene chloride (15 mL). After stirring for 1 h at room temperature, the solvent was evaporated in vacuo at 0-5 °C, and the crude acyl chloride was dissolved in the solvent of choice, preferably, methylene chloride (5 mL). This solution was added dropwise to a cooled (0 °C) mixture of the corresponding methyl glyoxalate imine (2 mmol) and triethylamine (0.63 mL, 4.5 mmol) in the same solvent (15 mL) during 10 min. After the ice bath was removed, the reaction mixture was heated at reflux for 20 h and then diluted in methylene chloride (50 mL) and washed successively with H_2O (50 mL), 1 M HCl (50 mL), saturated NaHCO₃ (50 mL), and H₂O (50 mL). Drying over MgSO₄ and evaporation of solvents yielded the corresponding crude β -lactams as mixtures of anti, cis and syn, cis isomers, which were separated by column chromatography (silica gel; eluent: hexane-methylene chloride 5:1).

Reaction between Copper β -Silyl Ester Enclates and Methyl Glyoxalate Imines. General Procedure for the Preparation of β -Lactams 29 and 30. A solution of (dimethylphenylsilyl)lithium (10 mL, 1 M in THF) was dropped over cold (0 °C) anhydrous copper(I) cyanide (0.45 g, 5 mmol) kept under nitrogen, and the mixture was stirred for 20 min at the same temperature. A solution of the corresponding α,β -unsaturated ester (5 mmol) in THF (10 mL) was dropwise added to the former suspension and the mixture was stirred for 20 min at 0 °C. Then, a solution of N-[(methoxycarbonyl)methylene]-p-anisidine (0.97 g, 5 mmol) in THF (10 mL) was added and the mixture was stirred at room temperature for 3 h. After this time, the reaction mixture was diluted in methylene chloride (30 mL), washed with saturated ammonium chloride (50 mL) at 0 °C, filtered through a pad of Celite, dried, and evaporated. The crude β -lactams 29 and 30 were purified by column chromatography (silica gel; eluent:hexane-methylene chloride 5:1).

cis, anti-3-[1'-(Dimethylphenylsilyl)alkyl]-4-(methoxycarbonyl)azetidin-2-ones (39). A solution of ammonium cerium(IV) nitrate (12.00 g, 21.9 mmol) in acetonitrile (65.5 mL) and water (7.28 mL) was added dropwise to a cooled (-10 °C) solution of the β -lactam cis, anti-22 (7.3 mmol) in acetonitrile (49 mL) and water (5.4 mL) within 15 min. Then, the reaction mixture was stirred at -10 °C for 3 h. On completion, 2 M sodium hydroxyde (43.5 mL) was added, and stirring was maintained for 30 min at room temperature. The resulting mixture was filtered through Celite, neutralized with 1 M hydrochloric acid, and extracted with methylene chloride $(3 \times 20 \text{ mL})$. After drying and evaporation of the solvents, the resulting crude was purified by column chromatography (silica gel; eluent:hexane-methylene chloride 1:1), vielding cis, anti-3-[1'-(dimethylphenylsilyl)ethyl]-4-(methoxycarbonyl)azetidin-2-one (39a) (1.70 g, 80%). Syrup. ¹H NMR (CDCl₃): δ 7.57 (m, 2 H, Ar), 7.36 (m, 2 H, Ar), 6.45 (s, 1 H, NH), 4.27 (d, 1 H, J = 5.4 Hz, CHCO₂Me), 3.76 (s, 3 H, OCH₃), 3.40 (dd, 1 H, J = 5.4 Hz, J = 11.7 Hz, CHCO), 1.26 (m, 1 H, CHSi),0.81 (d, 3 H, J = 7.2 Hz, CHCH₃), 0.45 (s, 3 H, SiCH₃), 0.39 (s, 3 H, SiCH₃). ¹³C NMR (CDCl₃): δ 173.3, 170.1, 134.3, 129.0, 127.6, 117.4, 61.2, 53.4, 52.3, 17.8, 14.0, -3.1, -4.2. MS: m/e 292 (M⁺). Similarly, cis, anti-23 (4.29 g, 7.3 mmol) afforded cis, anti-3- $[\alpha$ -(dimethylphenylsilyl)benzyl]-4-(methoxycarbonyl)azetidin-2-one (39b) (2.12 g, 82%). Mp: 106-107 °C (hexane/methylene chloride). ¹H NMR (CDCl₃): δ 7.31 (m, 4 H, Ar), 7.07 (m, 4 H, Ar), 6.67 (d, 2 H, Ar), 5.98 (s, 1 H, NH), 3.95 (d, 1 H, J = 4.5 Hz, CHCO₂Me), 3.74 (dd, 1 H, J = 4.5 Hz, J = 12.0 Hz, CHCO), 3.08 (s, 3 H, OCH₃), 2.72 (d, 1 H, J = 12 Hz, CHSi), 0.48 (s, 3 H, SiCH₃), 0.07 (s, 3 H, SiCH₃). ¹³C NMR (CDCl₃): δ 171.2, 168.8, 139.9, 134.9, 129.2, 128.0, 127.5, 125.2, 58.3, 52.1, 51.9, 33.1, -2.9, -3.1. MS: m/e 340 (M⁺ – 15). Anal. Calcd for C₂₀H₂₃NO₃Si: C, 67.95; H, 6.56; N, 3.96. Found: C, 67.53; H, 6.38; N, 3.78.

trans.anti-3-[1'-(Dimethylphenylsilyl)alkyl]-4-(methoxycarbonyl)azetidin-2-ones (43). A solution of ammonium cerium(IV) nitrate (6.56 g, 12.0 mmol) in water (42 mL) was added dropwise to a cooled (0 °C) solution of the β -lactam trans, anti-29a (1.59 g, 4.0 mmol) in acetonitrile (50 mL) within 15 min. Then, the reaction mixture was stirred at 0 °C for 30 min. On completion, the reaction mixture was taken up over water (160 mL) and extracted with ethyl acetate $(3 \times 60 \text{ mL})$. The organic layer was washed successively with saturated sodium hydrogen carbonate (140 mL), sodium hydrogen sulfite (4×100 mL), sodium hydrogen carbonate (30 mL), and brine (30 mL). After drying and evaporation of the solvents, the resulting crude was purified by column chromatography (silica gel; eluent:hexane-methylene chloride 1:1), yielding trans, anti-3-[1'-(dimethylphenylsilyl)alkyl]-4-(methoxycarbonyl)azetidin-2-one (43a) (1.34 g, 92%). Mp: 114-116 °C (Et₂O). ¹H NMR (CDCl₃): δ 7.53-7.36 (m, 5 H, Ar), $6.21 (s_b, 1 H, NH), 3.91 (d, 1 H, J = 2.4 Hz, CHCO_2Me), 3.74 (s,)$ 3 H, OCH_3), 3.34 (dd, 1 H, J = 2.4 Hz, J = 4.8 Hz, CHCO), 1.58-1.54 (m, 1 H, CHSi), 1.05 (d, 3 H, J = 7.5 Hz, CHCH₃), 0.38(s, 3 H, SiCH₃), 0.37 (s, 3 H, SiCH₃). ¹³C NMR (CDCl₃): δ 171.8, 169.9, 136.8, 133.9, 129.3, 127.3, 60.1, 52.4, 50.8, 18.4, 10.3, -4.7. MS: m/e 292 (M⁺). Anal. Calcd for C₁₅H₂₀NO₃Si: C, 62.01; H, 6.95; N, 4.82. Found: C, 62.10; H, 7.00; N, 4.85. The following compounds were obtained according to a similar procedure: $trans, anti-3-[\alpha-(dimethylphenylsilyl)benzyl]-4-(methoxy$ carbonyl)azetidin-2-one (43b) (1.15 g, 65%). Mp: 100-102 °C (hexane/methylene chloride). ¹H NMR (CDCl₃): δ 7.48-6.91 (m, 10 H, Ar), 6.20 (s_{b} , 1 H, NH), 3.65 (d, 1 H, J = 1.0 Hz, CHCO₂Me), $3.58 (dd, 1 H, J = 1.0 Hz, J = 10.6 Hz, CHCO), 3.55 (s, 3 H, OCH_2),$ 2.69 (d, 1 H, J = 10.3 Hz, CHPh), 0.41 (s, 3 H, SiCH₃), 0.26 (s, 3 H, SiCH₃). ¹³C NMR (CDCl₃): δ 171.5, 169.0, 140.3, 136.0, 129.3,

⁽³⁵⁾ For methods to prepare enantiomerically pure β -silyl carboxylic acids, see: (a) Fleming, I.; Kindon, N. D. J. Chem. Soc., Chem. Commun. 1987, 1177. (b) Fleming, I.; Lawrence, N. J. Tetrahedron Lett. 1990, 31, 3645. (c) Oppolzer, W.; Mills, R. J.; Pachinger, W.; Stevenson, T. Helv. Chim. Acta 1986, 69, 1542.

128.2, 127.6, 125.6, 59.9, 53.7, 52.2, 36.5, -2.9, -3.6. MS: m/e 340 (M⁺ - 15). Anal. Calcd for $C_{20}H_{23}NO_3Si$: C, 67.95; H, 6.56; N, 3.96. Found: C, 68.12; H, 6.59; N, 3.99. trans, anti-3-[1'-(Di-methylphenylsilyl)-2'-propyl]-4-(methoxycarbonyl)azetidin-2-one (43c) (1.07 g, 70%). Syrup. ¹H NMR (CDCl₃): δ 7.55-7.52 (m, 2 H, Ar), 7.38-7.33 (m, 3 H, Ar), 6.24 (s_b, 1 H, NH), 3.90 (d, 1 H, J = 2.7 Hz, $CHCO_2Me$), 3.73 (s, 3 H, OCH₃), 3.24 (t, 1 H, J = 2.7 Hz, J = 2.7 Hz, CHCO), 1.11 (s, 3 H, CCH₃), 1.01 (s, 3 H, CCH₃), 0.42 (s, 3 H, SiCH₃), 0.38 (s, 3 H, SiCH₃). ¹³C NMR (CDCl₃): δ 172.1, 169.2, 136.2, 134.7, 129.2, 127.6, 65.8, 52.4, 49.5, 22.7, 20.9, 20.5, -5.3, -5.4. MS: m/e 290 (M⁺).

Preparation of Dimethyl N-Benzoyl- β -[(dimethylphenylsilyl)alkyl]aspartates 41 and 45 from 39 and 43. General Procedure. Chlorotrimethylsilane (2.50 mL, 20 mmol) was added to a solution of 3-[1'-(dimethylphenylsilyl)alkyl]-4-(methoxycarbonyl)azetidin-2-one 39 or 43 (4.0 mmol) in methanol (10.0 mL) and the mixture was stirred for 4-10 h at room temperature (for 43c the reaction was refluxed for 4 h). On completion, the reaction mixture was evaporated, diluted with methylene chloride (30 mL), and washed with saturated sodium hydrogen carbonate (20 mL). Drying and evaporation of the solvents yielded crude dimethyl β -[(dimethylphenylsilyl)alkyl]aspartates 40 and 44, which were directly dissolved in methylene chloride (30 mL) and triethylamine (1.10 mL, 8 mmol). Benzoyl chloride (0.51 mL, 4.4 mmol) in methylene chloride (11 mL) was dropped over the preceding solution cooled to 0 °C within 10 min. and the resulting solution was stirred at room temperature for 4 h and then washed successively with water (10 mL), 1 M hydrochloric acid (10 mL), and saturated sodium hydrogen carbonate (10 mL). Drying and evaporation of the solvents yielded crude dimethyl N-benzoyl- β -[(dimethylphenylsilyl)alkyl]aspartates 41 and 45, which were purified by column chromatography (silica gel; eluent:methylene chloride-hexane (1:15).

Preparation of 3-(1'-Hydroxyalkyl)-4-(methoxycarbonyl) β -Lactams 17, 33, and 34 and Dimethyl N-Benzoyl- β -(hydroxyalkyl)aspartates 42 and 46. General Procedure. HBF₄·OEt₂ (1.06 mL, 10 mmol) was added to a cooled (0 °C) solution of 3-(1'-dimethylphenylsilylalkyl)-4-(methoxycarbonyl)azetidin-2-one (5 mmol) or dimethyl N-benzovl B-[(dimethylphenylsilyl)alkyl]aspartate (5 mmol) in methylene chloride (20 mL) and the mixture was stirred at room temperature for 1 h. After this time, the reaction mixture was washed with cold water (10 mL) and brine (20 mL). The organic solvent was dried and evaporated to yield the corresponding dimethylfluorosilane. The crude product was dissolved in cold (0 °C) 32% peracetic acid in acetic acid (25 mL, 25 mmol), and triethylamine (1.25 mL, 11 mmol) was added dropwise within 5 min at the same temperature. The mixture was stirred at room temperature for 3 h, then methylene chloride (30 mL) was added, and the resulting solution was successively washed with 1 M HCl (30 mL), 40% NaHSO₃ (30 mL), NaHCO₃ (30 mL), and water (30 mL). Drying and evaporation afforded the hydroxy derivatives 33 and 34 or 42 and 46. The crude products were purified by column chromatography (silica gel, eluent:methylene chloride:hexane 1:3). anti,trans-3-(1'-Hydroxyethyl)-4-(methoxycarbonyl)-1-(p-methoxyphenyl)azetidin-2-one (33) (1.32 g, 95%). Mp: 91–92 °C (hexane/methylene chloride). ¹H NMR (CDCl₃): δ 7.25 (d, 2 H, Ar), 6.86 (d, 2 H, Ar), 4.42 (d, 1 H, J = 2.4 Hz, $CHCO_2Me$), 4.24-4.20 (m, 1 H, CHOH), 3.80 (s, 3 H, OCH₃), 3.78 (s, 3 H, OCH_3), 3.38 (dd, 1 H, J = 2.4 Hz, J = 5.4 Hz, CHCO), 1.43 (d, 3 H, J = 6.6 Hz, CH₃). ¹³C NMR (CDCl₃): δ 170.3, 163.7, 156.5, 129.3, 117.9, 114.4, 65.7, 61.2, 55.5, 53.6, 52.8, 21.2. Anal. Calcd for C14H17NO5: C, 60.19; H, 6.15; N, 5.01. Found: C, 60.20; H, 6.15; N, 5.03. anti, trans-3-(a-Hydroxybenzyl)-4-(methoxycarbonyl)-1-(p-methoxyphenyl)azetidin-2-one (34) (1.02 g, 60%). Syrup. ¹H NMR (CDCl₃): δ 7.52-7.18 (m, 5 H, Ar), 7.21 (d, 2 H, Ar), 6.84 (d, 2 H, Ar), 5.13 (d, 1 H, J = 5.24 Hz, CHOH), 4.40 $(d, 1 H, J = 2.6 Hz, CHCO_2Me), 3.76 (s, 3 H, OCH_3), 3.74 (dd, J)$ 1 H, J = 2.6 Hz, J = 5.4 Hz, CHCO), 3.72 (s, 3 H, OCH₃). ¹³C NMR (CDCl₃): δ 170.1, 163.1, 156.5, 140.0, 130.6, 128.9, 128.7, 128.2, 126.6, 118.0, 114.4, 71.9, 61.1, 55.4, 54.0, 52.8. Anal. Calcd for C₁₉H₁₉NO₅: C, 66.85; H, 5.61; N, 4.10. Found: C, 66.91; H, 5.62; N, 4.14. Methyl $2(S^*)$ -(benzoylamino)- $4(R^*)$ -hydroxy-3-(S*)-(methoxycarbonyl)pentanoate (42a) (1.11 g, 72%). Syrup. ¹H NMR (CDCl₃): δ 7.80 (d, 2 H, Ar), 7.52 (m, 3 H, Ar), 5.28 (dd, 1 H, J = 5.4 Hz, J = 8.6 Hz, CHNH), 4.09 (m, 1 H, CHOH), 3.80

 $(s, 3 H, OCH_3), 3.76 (s, 3 H, OCH_3), 3.04 (dd, 1 H, J = 2.7 Hz,$ J = 5.4 Hz, CHCO), 1.37 (d, 3 H, J = 6.6 Hz, CH₃CH). ¹³C NMR $(CDCl_3): \delta 171.9, 171.0, 167.4, 132.3, 132.0, 128.6, 127.5, 66.7, 53.6,$ 53.0, 52.3, 52.2, 22.0. MS: m/e 279 (M⁺ - 30). Methyl 2(S^{*})- $(benzoylamino)-4(R^*)-hydroxy-3(S^*)-(methoxycarbonyl)-4$ phenylbutanoate (42b) (1.21 g, 65%). Syrup. ¹H NMR (CDCl₃): δ 7.80 (d, 2 H, Ar), 7.53 (m, 2 H, Ar), 7.47 (d, 2 H, Ar), 7.38 (d, 4 H, Ar), 5.18 (dd, 1 H, J = 5.1 Hz, J = 8.1 Hz, CHNH), 5.09 (d, 1 H, J = 5.1 Hz, CHOH, 3.79 (s, 3 H, OCH₃), 3.66 (s, 3 H, OCH₃), 3.53 (t, 1 H, J = 5.1 Hz, CHCO). ¹³C NMR (CDCl₃): δ 172.5, 171.2, 167.8, 141.1, 132.6, 129.2, 129.0, 128.5, 127.7, 126.4, 72.7, 53.8, 53.6, 53.3, 52.9. MS: m/e 237 (M⁺ - 144). Methyl 2(S^{*})-(benzovlamino)-4(S^*)-hydroxy-3(R^*)-(methoxycarbonyl)pentanoate (46a) (1.08 g, 70%). Mp: 200-202 °C (methanol). ¹H NMR (CDCl_a); δ 7.92-7.90 (m, 2 H, Ar), 7.55-7.41 (m, 3 H, Ar), 5.08 (qd, 1 H, J = 6.7 Hz, J = 8.9 Hz, CHOH), 4.93 (dd, 1 H, J = 8.9 Hz, J =7.5 Hz, CHNH), 4.02 (t, 1 H, J = 8.9 Hz, CHCO₂Me), 3.76 (s, 3 H, OCH₃), 3.40 (s, 3 H, OCH₃), 1.34 (d, 3 H, J = 6.6 Hz, CH₃). ¹³C NMR (CDCl₃): δ 172.4, 169.6, 166.8, 132.6, 131.5, 127.9, 127.2, 74.5. 72.9, 52.1, 50.7, 47.9, 17.0. Anal. Calcd for C₁₅H₁₉NO₆: C, 58.25; H, 6.20; N, 4.53. Found: C, 59.02; H, 6.25; N, 4.56. Methyl $2(S^*)$ -(benzoylamino)-4(S^*)-hydroxy-3(R^*)-(methoxycarbonyl)-4-phenylbutanoate (46b) (1.67 g, 90%). Syrup. ¹H NMR (CDCl₃): δ 7.8–7.24 (m, 2 H, Ar), 5.01 (d, 1 H, J = 8.6 Hz, CHOH), 4.69 $(dd, 1 H, J = 3.1 Hz, J = 8.6 Hz, CHNH), 3.77 (s, 3 H, OCH_3),$ 3.67 (s, 3 H, OCH₃), 3.50 (dd, 1 H, J = 3.1 Hz, J = 8.6 Hz, CHCO₂Me). MS: m/e 265 (M⁺ - 106). Anal. Calcd for C20H21NO6: C, 64.68; H, 5.70; N, 3.77. Found: C, 64.74; H, 5.71; N, 3.81. Methyl 2(S*)-(benzoylamino)-4-hydroxy-3(R*)-(methoxycarbonyl)-4-methylpentanoate (46c) (1.45 g, 90%). Syrup. ¹H NMR (CDCl₃): δ 8.13–7.61 (m, 5 H, Ar), 5.20 (dd, 1 H, J = 2.2Hz, J = 8.2 Hz, CHNH), 4.06 (s, 3 H, OCH₃), 4.03 (s, 3 H, OCH₃), 3.46 (d, 1 H, J = 2.2 Hz, CHCO₂Me), 1.18 (s, 3 H, CH₂). MS: m/e265 (M⁺ - 44). Anal. Calcd for C₁₆H₂₁NO₆: C, 59.43; H, 6.55; N, 4.33. Found: C, 59.22; H, 6.50; N, 4.29.

When the anti-3-[1'-(dimethylphenylsilyl)ethyl]-4-(methoxycarbonyl)-1-(p-methoxyphenyl)azetidin-2-one (15a) (1.98 g. 5 mmol) was subjected to the above protocol, a mixture of compounds anti-17 and 18 was obtained, which was separated by column chromatography (silica gel, eluent:methylene chloride: hexane 1:8), yielding compound anti-17 (0.56 g, 40%) [Mp: 165–167 °C (hexane-methylene chloride). ¹H NMR (CDCl₃): δ 7.27 (d, 2 H, Ar), 6.86 (d, 2 H, Ar), 4.59 (d, 1 H, J = 6.3 Hz, $CHCO_2Me$), 4.18 (q, 1 H, J = 6.3 Hz, CHOH), 3.78 (s, 3 H, OCH_3), $3.77 (s, 3 H, OCH_3), 3.60 (t, 1 H, J = 6.3 Hz, CHCO), 1.38 (d, 3 Hz)$ H, J = 6.3 Hz, CH_3 CH). ¹³C NMR (CDCl₃): δ 170.3, 163.7, 156.5, 130.7, 117.9, 114.4, 65.7, 61.2, 55.5, 53.6, 52.8. Anal. Calcd for C₁₄H₁₇NO₅: C, 60.19; H, 6.15; N, 5.01. Found: C, 60.29; H, 6.31; N, 5.22.] and the lactone 18 (0.37 g, 30%) [Mp: 162 °C (hexane-methylene chloride). ¹H NMR (CDCl₃): δ 7.51 (d, 2 H, Ar), 6.89 (d, 2 H, Ar), 4.84 (dq, 1 H, J = 6.6 Hz, J = 7.8 Hz, CHOCO),4.62 (d, 1 H, J = 4.8 Hz, CHN), 4.05 (dd, 1 H, J = 4.8 Hz, J =7.8 Hz, CHCO), 3.79 (s, 3 H, OCH₃), 1.65 (d, 3 H, J = 6.6 Hz, CH_3). ¹³C NMR (CDCl₃): δ 170.4, 160.7, 156.7, 130.5, 118.3, 114.4, 75.1, 55.5, 54.5, 53.7, 17.9. MS: m/e 247 (M⁺). Anal. Calcd for C₁₃H₁₃NO₄: C, 63.15; H, 5.31; N, 5.66. Found: C, 63.17; H, 5.10; N. 5.48.1

6-(p-Methoxyphenyl)-2-methyl-3-oxa-6-azabicyclo[3.2.0]heptane-4,7-dione (19). A solution of the alcohol syn-17 (0.56 g, 2 mmol) and boron trifluoride etherate (0.05 mL, cat.) in methylene chloride (10 mL) was stirred at room temperature for 20 min. The reaction mixture was washed with water (20 mL) and brine (20 mL), dried, and evaporated, affording the lactone 19 (0.39 g, 80%). Mp: 163-165 °C (hexane-methylene chloride). ¹H NMR (CDCl₃): δ 7.49 (d, 2 H, Ar), 6.90 (d, 2 H, Ar), 5.08 (dq, 1 H, J = 6.6 Hz, J = 1.8 Hz, CHOCO), 4.67 (d, 1 H, J = 4.5 Hz, CHN), 3.79 (s, 3 H, OCH₃), 3.73 (dd, 1 H, J = 1.8 Hz, J = 4.5 Hz, CHCO), 1.50 (d, 3 H, J = 6.6 Hz, CH_3). ¹³C NMR (CDCl₃): δ 170.4, 162.9, 157.3, 130.9, 118.3, 114.5, 75.2, 55.8, 55.5, 52.2 a.8. MS: m/e 247 (M⁺). Anal. Calcd for C₁₃H₁₃NO₄: C, 63.15; H, 5.31; N, 5.66. Found: C, 63.11; H, 5.12; N, 5.44.

Acknowledgment. The present work has been supported by Gobierno Vasco (Project 170-215-0017/89) and, in part, by Diputación Foral de Gipuzkoa (Project BOG-85/10.05/90). Two grants from the Ministerio de

Educación y Ciencia to R.U. and M.I. are gratefully acknowledged.

Registry No. 3, 65750-57-4; 8, 18707-60-3; 9, 103-26-4; 10a, 138234-97-6; 10b. 138234-98-7; 10c. 138234-99-8; 10d. 138235-00-4; 11a, 138235-01-5; 11b, 138235-02-6; 11c, 138235-03-7; 14, 72079-55-1; anti-15a, 128474-84-0; syn-15a, 128571-95-9; anti-15b, 138235-04-8; syn-15b, 138331-54-1; anti-15c, 138235-05-9; syn-15c, 138331-55-2; anti-15d, 138235-06-0; syn-15d, 138331-56-3; anti-17, 135683-14-6; syn-17, 135758-02-0; 18, 138235-07-1; 19, 138331-57-4; (E)-20, 131533-35-2; (Z)-20, 131533-34-1; 21, 138235-08-2; anti-22, 138235-09-3; syn-22, 138331-58-5; anti-23, 138235-10-6; 29a, 128571-96-0; 29b, 131533-36-3; 29c, 131533-38-5; 29d, 131533-37-4; 30b, 131614-09-0; 30d, 138331-59-6; 33, 131533-32-9; 34, 13823511-7; 35, 131614-08-9; 37, 131533-33-0; 38, 138235-12-8; 39a, 138235-13-9; 39b, 138235-14-0; 41a, 138235-15-1; 41b, 138235-16-2; 42a, 138235-17-3; 42b, 138235-18-4; 43a, 138331-60-9; 43b, 138331-61-0; 43c, 138235-19-5; 45a, 138331-62-1; 45b, 138331-63-2; 45c, 138235-20-8; 46a, 138331-64-3; 46b, 138331-65-4; 46c, 138235-21-9.

Supplementary Material Available: Characterization data for 10a, 10b, 10d, 11a, 11b, 11c, anti-15a, syn-15a, anti-15b, syn-15b, anti-15c, syn-15c, anti-15d, syn-15d, anti-22, syn-22, 23, 29a, 29b, 29c, 29d, 37, 38, 41a, 41b, 45a, 45b, and 45c (6 pages). This material is contained in many libraries on microfiche, immediately follows this article in the microfilm version of the journal, and can be ordered from the ACS; see any current masthead page for ordering information.

Reaction of Nucleic Acid Bases with α -Acetylenic Esters. 5.¹ Synthesis and **Properties of Adenosine and Cytidine Derivatives**

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 α -Acetylenic esters are able to react under mild experimental conditions with the base moiety of adenosine and cytidine, while guanosine is unreactive. A double reaction of the triple bond and the ester group of the reagent with the NH₂ group and the adjacent ring nitrogen of the base yields derivatives in which an additional pyrimidone ring is fused to the original base. These derivatives can exist in two isomeric forms. In alkaline solution, or by prolonged heating in water, the medium pyrimidine ring of adenosine derivatives opens by loss of carbon 5. If the derivatization is performed with chlorotetrolic (4-chloro-2-butynoic) acid esters, the modified nucleobases contain a chloromethyl side chain. Tests of the alkylating abilities of the latter in the two isomeric adenosine derivatives show that the chlorine can be easily substituted by a thiol in the presence of alkali; a partial Dimroth rearrangement of one of the reaction products is observed. The reaction with amines is accompanied by ring opening. Nucleic acids containing these alkylating base derivatives can be cross-linked to other macromolecules such as solid supports or contact proteins.

Introduction

Nucleic acid bases having an exocyclic NH₂ and an adjacent ring nitrogen are able to react with certain types of electrophilic bifunctional reagents, yielding derivatives in which an additional five- or six-membered heterocycle is fused to the original purine or pyrimidine. Many of these reagents contain a halogen atom and/or an unsaturated group including C=O,³⁻⁸ C=C,⁹⁻¹² C=N,¹³⁻¹⁶ and

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- 1969, 8, 238. (4) Moschel, R. C.; Leonard, N. J. J. Org. Chem. 1976, 41, 294.
 (5) Kochetkov, N. K.; Shibaev, V. N.; Kost, A. A. Tetrahedron Lett.
- 1971. 1993.

- (6) (a) Barrio, J. R.; Secrist, J. A.; Leonard, N. J. Biochem. Biophys.
 Res. Commun. 1972, 46, 597. (b) Kayasuga-Mikado, K.; Hashimoto, T.;
 Negishi, T.; Negishi, K.; Hayatsu, H. Chem. Pharm. Bull. 1980, 28, 932.
 (7) Sattsangi, P. D.; Leonard, N. J.; Frihart, C. R. J. Org. Chem. 1977, 42, 3292.
- (8) Leonard, N. J.; Cruickshank, K. A. J. Org. Chem. 1985, 50, 2480.
 (9) (a) Chung, F. L.; Young, R.; Hecht, S. S. Cancer Res. 1984, 44, 990.
 (b) Chung, F. L.; Roy, K. R.; Hecht, S. S. J. Org. Chem. 1988, 53, 14.
 (10) Meerman, J. H. N.; Pearson, P. G.; Meier, G. P.; Nelson, S. D. J.
 Org. Chem. 1988, 53, 30.
 (11) Neudecker, T.; Lutz, D.; Eder, E.; Henschler, D. Mutat. Res. 1981, 41, 27
- 91, 27

(12) Laib, R. J.; Bolt, H. M. Arch. Toxicol. 1978, 39, 235.

N=C=O.¹⁷⁻¹⁹ The long known chlorotetrolic (4-chloro-2-butynoic) acid esters, ClCH₂C=CCOOR,²⁰⁻²² have such functions and therefore, like other bifunctional nucleobase reagents, should be able to react with the amidine -N== $C(NH_2)$ - system of adenine or cytosine or the guanidine $-N=C(NH_2)NH$ - system of guanine. This expectation was strengthened by our earlier works which have shown that methyl chlorotetrolate can be used as a bifunctional protein modifier, reacting through its chloromethyl group and the triple bond with protein nucleophiles such as amine, thiol, or imidazole.²³ A similar behavior toward

- (13) Furukawa, Y.; Miyashita, O.; Honjo, M. Chem. Pharm. Bull. 1974, 22, 2552
- (14) Hosmane, R. S.; Leonard, N. J. J. Org. Chem. 1981, 46, 1457.
 (15) Agasimundin, Y. S.; Oakes, F. T.; Kostuba, L. J.; Leonard, N. J.
 J. Org. Chem. 1985, 50, 2468; J. Am. Chem. Soc. 1984, 106, 6847.
 (16) Agasimundin, Y. S.; Oakes, F. T.; Leonard, N. J. J. Org. Chem.

- 1985, 50, 2474.
 (17) Kumar, S.; Leonard, N. J. J. Org. Chem. 1988, 53, 3959.
 (18) (a) Camus, P. Doctorate Thesis, University of Grenoble, 1988. (b)
 Camus, P.; Lhomme, M. F.; Lhomme, J. Tetrahedron Lett. 1989, 30, 467.
- (19) Nair, V.; Turner, G. A. Tetrahedron Lett. 1984, 25, 247. (20) (a) Olomucki, M. C. R. Acad. Sci. 1958, 246, 1877. (b) Olomucki,
- M.; Marszak, I. Bull. Soc. Chim. Fr. 1959, 315. (c) Angew. Chem. 1959, 71, 314. (d) Olomucki, M. Ann. Chim. (Paris) 1960, 5, 845.
- (21) Olomucki, M.; Le Gall, J. Y.; Barrand, I. J. Chem. Soc., Chem. Commun. 1982, 1290.
- (22) Olomucki, M.; Le Gall, J. Y. Org. Synth. 1987, 65, 47.
 (23) (a) Olomucki, M.; Diopoh, J. C. R. Acad. Sci., Ser. D. 1977, 284,
 2293. (b) Diopoh, J.; Olomucki, M. Bioorg. Chem. 1982, 11, 463.